

# Consequences of Self and Foreign Superantigen Interaction with Specific $V_{\beta}$ Elements of the Murine $TCR\alpha\beta$

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The  $\alpha\beta$  T-cell receptor ( $TCR\alpha\beta$ ) recognizes a ligand composed of an antigen fragment complexed with a product of the major histocompatibility complex (MHC). The repertoire of receptors is limited both by the germ line of receptor variable elements and by selective events that take place during T-cell development. The current view is that the germ-line repertoire is expressed in the thymus randomly but that only those T cells bearing receptors that successfully interact with MHC molecules expressed on thymic cortical epithelial cells are allowed to mature (Bevan and Fink 1978; Zinkernagel et al. 1978; Kisielow et al. 1988; Sha et al. 1988). Furthermore, during the process of establishing tolerance to self-antigens, this positively selected population is further reduced by the deletion or inactivation of clones whose receptors continue to interact with self-antigen/MHC ligands (Kappler et al. 1987a, 1988; MacDonald et al. 1988; Pullen et al. 1988). Thus, positive and negative selections reduce expressed receptor repertoire in the periphery to a fraction of the germ-line repertoire.

Five variable elements contribute to the specificity of the  $TCR\alpha\beta$ :  $V_{\alpha}$  (variable),  $J_{\alpha}$  (joining),  $V_{\beta}$ ,  $D_{\beta}$  (diversity), and  $J_{\beta}$ . Despite the fact that in most circumstances all five components together determine receptor specificity, a study of the variation of expression of individual  $V_{\beta}$  elements in different mouse strains has produced a wealth of information on genetic factors effecting the  $TCR\alpha\beta$  repertoire. Several studies have shown the influence of thymic-positive selection on expression of particular  $V_{\beta}$  elements (Blackman et al. 1989; Palmer et al., this volume). More striking are the many mechanisms in mice that prevent the expression of particular  $V_{\beta}$  elements in individual mice. Some of these are genetic defects within the  $V_{\beta}$  complex (Chou et al. 1987a; Kappler et al. 1987b; Wade et al. 1988). However, the most surprising mechanism limiting  $V_{\beta}$  expression involves the induction of tolerance to a set of unusual self-antigens, which we have termed "superantigens" (White et al. 1989). Superantigens combine with MHC molecules to generate ligands that are recognized by all  $TCR\alpha\beta$ s bearing one or a few  $V_{\beta}$  elements with little input from the other receptor vari-

able elements (Kappler et al. 1987a, 1988; MacDonald et al. 1988; Pullen et al. 1988). The consequence is that mice carrying these antigens delete nearly all T cells bearing the relevant  $V_{\beta}$  elements while establishing self-tolerance. Since there are only a total of 22 different  $V_{\beta}$  elements in the mouse, these self-superantigens drastically alter the repertoire of the mice in which they occur.

Superantigens have also been identified as products of microorganisms. Particularly striking examples are the toxins of *Staphylococcus aureus*, which cause a number of diseases in humans and other animals (Bergdoll 1983). These toxins have been recognized as powerful T-cell stimulants for many years, but the role of T cells in their mode of action has not been pursued. Recently, we and other investigators have shown that these toxins are superantigens in that each stimulates a set of human or mouse T cells bearing particular  $V_{\beta}$  elements (Choi et al. 1989; Janeway et al. 1989; Kappler et al. 1989b; White et al. 1989).

In this paper, we review some of the properties of the bacterial and the polymorphic murine superantigens and attempt to tackle the question of their possible function in bacteria and mice. We present evidence that T-cell stimulation by bacterial superantigens is critical to their toxic properties. In addition, we demonstrate that in mice widespread polymorphisms in superantigens as well as in  $V_{\beta}$  structural genes results in a state where the mouse population as a whole expresses all  $V_{\beta}$  elements but that the number of  $V_{\beta}$  elements expressed in an individual mouse is generally quite limited. We propose that this state may be an evolutionary equilibrium balancing the advantage of a large, diverse T-cell repertoire against the disadvantage of possessing too many potential targets for the microbial superantigen toxins.

## RESULTS

### Germ-line Limitations on $V_{\beta}$ Expression

So far, 22 functional murine  $V_{\beta}$  elements have been described (Fig. 1). However, no mouse has been found

|   |   |
|---|---|
| -2--4-16-10-1---5 <sup>2</sup> -8 <sup>3</sup> -5 <sup>1</sup> -8 <sup>2</sup> -8 <sup>1</sup> -13-12-11-9--6--15---19-17-3---7--18--DJC $\beta$ 1-DJC $\beta$ 2--14- | BALB/c, B6<br>DBA, C3H, etc.                                  |
|   | x x   |
| -2--4-16-10-1-...   | ...-6--15---19-17-3---7--18--DJC $\beta$ 1-DJC $\beta$ 2--14- |
|   | SJL, SWR,<br>C57L, C57BR                                      |
| -2--4-16-10-1-...   | ...-3---7--18--DJC $\beta$ 1-DJC $\beta$ 2--14-               |
|   | RIII  |
| -2--4-16-10-1---5 <sup>2</sup> -8 <sup>3</sup> -5 <sup>1</sup> -8 <sup>2</sup> -8 <sup>1</sup> -13-12-11-9--6--15---19-17-3---7--18--DJC $\beta$ 1-DJC $\beta$ 2--14- | Florida<br>Wild Mice  |
|   | ? x   |
| -2--4-16-10-1-...   | ...---19-17-3---7--18--DJC $\beta$ 1-DJC $\beta$ 2--14-       |
|   | Florida<br>Wild Mice  |
|   | ? x   |

**Figure 1.** Deletions and mutations within the mouse  $V_{\beta}$  complex. The genomic organization of the mouse  $V_{\beta}$  complex is shown for various laboratory strains and for wild mice from central Florida. Approximate position of deletions is shown. (X)  $V_{\beta}$  elements known to carry point mutations that inactivate the gene. Data are from this paper and Behlke et al. (1986), Malissen et al. (1986), Kappler et al. (1987b), Chou et al. (1987a,b), Lai et al. (1987, 1988), Wade et al. (1988), and Haqqi et al. (1989). Data from  $V_{\beta}$  19 was from D. Loh (pers. comm.).

that carries all 22 functional  $V_{\beta}$  genes because various deletions and mutations eliminate at least some  $V_{\beta}$  elements in all mice examined thus far. Three  $V_{\beta}$  haplotypes have been identified in laboratory mice. The haplotype carried by most strains has a full complement of  $V_{\beta}$  genes; however, the  $V_{\beta}$  17 gene carries a single-base mutation generating a termination codon near the 3' end of the coding region (Wade et al. 1988), and the  $V_{\beta}$  19 gene is inactivated by a frameshift caused by a single-base deletion in the leader (D. Loh, pers. comm.). Loh and colleagues have described a large deletion in SJL and a few other strains of mice in the middle of the  $V_{\beta}$  complex, that eliminates nine functional  $V_{\beta}$  elements (Chou et al. 1987a). These strains have a functional  $V_{\beta}$  17 and  $V_{\beta}$  19 gene. Recently, a similar but more extensive deletion has been found in another laboratory strain, RIII (Haqqi et al. 1989).

Such massive elimination of functional  $V_{\beta}$  elements should be disadvantageous to mice since the mice carrying these deletions should have a substantially reduced TCR $\alpha\beta$  repertoire. Therefore, one could expect that in the wild the pressure of dealing with environmental pathogens might select against mice carrying the mutations and deletions. On the contrary, Huppi et al. (1988) have found that a deletion similar to that of SJL mice is common in mice from Hebrides. We have examined the  $V_{\beta}$  complex of a series of 39 wild mice from central Florida. Southern blot analysis of the  $V_{\beta}$  17 gene in these mice showed a restriction-fragment-length polymorphism pattern identical to the inactive  $V_{\beta}$  17 allele (Kappler et al. 1987b; Wade et al. 1988), and no T cells were detected in these mice reactive with an antibody specific for  $V_{\beta}$  17. In addition, a deletion was found in the  $V_{\beta}$  complex in many of these mice. This deletion was intermediate in length between the two previously described deletions, involving 11  $V_{\beta}$  elements (Fig. 1), and was frequent enough in these populations so that 10 of the 39 mice examined were homozygous for the deletion and therefore lacked any

T cells bearing the deleted  $V_{\beta}$  elements (Table 2). Thus, for some reason, mice in the wild tolerate at high-frequency genetic mutations and deletions that limit the TCR $\alpha\beta$  repertoire in some mice.

#### Deletion of T Cells by Polymorphic $V_{\beta}$ -specific Self-superantigens

A similar picture of limited  $V_{\beta}$  expression has emerged during examination of the  $V_{\beta}$ -specific self-superantigens. We identified the first antigen of this type as a B-cell product that combined with the MHC class II molecule, IE, to form a ligand for virtually all T cells whose TCR $\alpha\beta$  contained  $V_{\beta}$  17a (Kappler et al. 1987b; Marrack and Kappler 1988). IE<sup>+</sup> mice carrying the functional  $V_{\beta}$  17a gene delete virtually all  $V_{\beta}$  17a<sup>+</sup> T cells while establishing self-tolerance (Kappler et al. 1987a, 1989a). Since then, we and other investigators have identified the  $V_{\beta}$  specificity of a number of other self-superantigens. For example, mice carrying the minor lymphocyte-stimulating (Mls-1<sup>a</sup>) antigen, long known as a powerful alloantigen, have been shown to eliminate virtually all T cells bearing  $V_{\beta}$  6 (MacDonald et al. 1988),  $V_{\beta}$  8.1 (Kappler et al. 1988), and  $V_{\beta}$  9 (Happ et al. 1989). Likewise, mice carrying the Mls-2<sup>a</sup> or Mls-3<sup>a</sup> antigen eliminate T cells bearing  $V_{\beta}$  3 (Pullen et al. 1988, 1989). Examples of these phenomena are shown in Figure 2 and 3, and the properties of the  $V_{\beta}$ -specific self-superantigens reported so far are summarized in Table 1.

Some of the most striking features of these antigens are the extensive polymorphisms controlling their expression. Thus, in every case, the superantigen and/or the MHC-restricting elements that present it are polymorphic, so that a functional ligand appears in only some strains of mice. For example, the superantigens that delete T cells bearing  $V_{\beta}$  17a,  $V_{\beta}$  5, or  $V_{\beta}$  11 all require an IE molecule for presentation. Therefore, even in the presence of the superantigen, IE<sup>-</sup> strains of





Table 2. High Frequency of V $\beta$  Elimination in Wild Mice

|   | Mechanism of V $\beta$ elimination               |                                |                                    |                      |                   |                   |
|---|--|--------------------------------|------------------------------------|----------------------|-------------------|-------------------|
|   | homozygous<br>gene<br>deletion                   | homozygous<br>gene<br>mutation | tolerance<br>to self-superantigens |                      |                   |                   |
|   |  |                                | Mls-1 <sup>a</sup>                 | Mls-2/3 <sup>a</sup> | IE <sup>+</sup> ? | IE <sup>+</sup> ? |
| V $\beta$ s<br>eliminated                           | 5.1 + 2<br>8.1 + 2 + 3<br>13, 12, 11<br>9, 6, 15 | 17                             | 6, 8.1<br>9<br>8.2                 | 3                    | 5.1 + 2           | 11                |
| Portion of<br>wild mice<br>with this<br>elimination | 10/39  | 39/39                          | 8/29                               | 32/39                | 21/29             | 2/29              |

toxic microbial products that are themselves V $\beta$ -specific superantigens and use this property in their mode of action.

A number of microbial products have been identified as potent T-cell stimulators in vitro in mouse and man. The related exotoxins of *S. aureus* are particularly good examples (Bergdoll 1983; Couch et al. 1988). In humans, these toxins are responsible for a large portion of food poisoning cases (staphylococcal enterotoxins A to E [SEA to SEE]), as well as other diseases such as scalded skin syndrome (exfoliating toxin) and toxic shock syndrome (toxic shock syndrome toxin [TSST-1]). Originally considered nonspecific mitogens, recent work has shown that these toxins must bind MHC molecules in order to stimulate T cells (Fischer et al. 1989; Fraser 1989) and that for each toxin only T cells bearing particular V $\beta$  elements respond (Choi et al. 1989; Janeway et al. 1989; Kappler et al. 1989b; White et al. 1989). Table 4 summarizes our results in mice using some of the *S. aureus* toxins. We found that each toxin stimulates a different set of V $\beta$ -bearing T cells. Even very closely related toxins (Couch et al. 1988), such as SEB and SEC1 or SEA and SEE, stimulate different T cells. Since most of the toxins stimulate T cells bearing any of several V $\beta$  elements, the proportion of total T cells stimulated is in some cases very high. For example, in the mouse SEB stimulates T cells bearing V $\beta$ 3, V $\beta$ 8, V $\beta$ 7, or V $\beta$ 17a. Depending on the strain of mouse, this can account for over 30% of all T cells. Similarly, in humans, we have found that these toxins stimulate T cells bearing particular V $\beta$  elements and that particular toxins often stimulate T cells bearing homologous V $\beta$  elements in mice and humans (Choi et al. 1989).

### T Cells Bearing Specific V $\beta$ s Required in the Mode of Action of SEB

Despite the considerable variability of the primary sequences of these toxins, their ability to act as superantigens appears to be a conserved property. Furthermore, since each has a different V $\beta$  specificity, their superantigen function appears not to be an accidental consequence of the conservation of some common structural element required for some other function. Therefore, it is reasonable to suppose that the pathology associated with these toxins (e.g., vomiting, diarrhea, skin rashes, and shock) may be an indirect consequence of the massive in vivo activation of T cells and release of powerful lymphokine mediators rather than a direct effect of the toxins on the target tissue. This idea is difficult to test directly in humans where most of the pathology associated with these toxins has been identified so we have begun to study the effects of these toxins in mice in vivo.

We have begun our experiments with SEB. Mice given a single injection of SEB develop a shock-like syndrome within 24 hours. Usually, high doses are required (> 50  $\mu$ g/mouse), correlating with the observation that this toxin, which was isolated from a strain of *Staphylococcus* that infects humans, must be used at a higher concentration in vitro to stimulate murine T cells than human T cells. The most obvious in vivo symptom in mice from this toxin is extremely rapid weight loss with wasting and in some cases death within 2–3 days. Those mice that survive for 3 days usually recover completely over the next few days. We have used this simple assay to assess the importance of the superantigen properties of SEB for its in vivo toxicity.

Table 3. Amino Acid Substitutions in the V $\beta$ 8.2 Gene of Wild Mice

| Mouse   | V $\beta$ | Mls-1 <sup>a</sup><br>reactive | Binds<br>MAb<br>F23.2 | Amino Acid |    |    |    |    |
|---------|-----------|--------------------------------|-----------------------|------------|----|----|----|----|
|         |           |                                |                       | 8          | 22 | 51 | 70 | 71 |
| C57BL/6 | 8.2       | –                              | +                     | N          | N  | G  | E  | N  |
| Wild    | 8.2       | +                              | –                     | S          | D  | D  | K  | E  |
| C57BL/6 | 8.1       | +                              | –                     | S          | H  | D  | E  | N  |

C57BL/6 sequences from Chou et al. (1987b). Amino acid numbering starts at the alanine at the amino termi of the mature proteins.

Table 4. *S. aureus* Toxins Are Superantigens in the Mouse

| <i>S. aureus</i><br>toxin | Response of T cells bearing $V_{\beta}$ |   |   |   |   |   |   |   |   |   |   |    |    |    |    |    |    |      |
|---------------------------|---|---|---|---|---|---|---|---|---|---|---|----|----|----|----|----|----|------|
|                           |   |   |   |   | 5 |   |   |   | 8 |   |   |    |    |    |    |    |    |      |
|                           | 1                                       | 2 | 3 | 4 | 1 | 2 | 6 | 7 | 1 | 2 | 3 | 10 | 11 | 12 | 14 | 15 | 16 | 17   |
| SEA                       | +                                       | - | + | - | - | - | - | - | - | - | - | -  | +  | -  | -  | -  | -  | +    |
| SEE                       | -                                       | - | - | - | - | - | - | - | - | - | - | -  | +  | -  | -  | +  | -  | n.d. |
| SEB                       | -                                       | - | + | - | - | - | - | + | + | + | + | -  | -  | -  | -  | -  | -  | +    |
| SEC1                      | -                                       | - | + | - | - | - | - | - | - | + | - | -  | -  | -  | -  | -  | -  | +    |
| TSST                      | -                                       | - | + | - | - | - | - | - | - | - | - | -  | -  | -  | -  | +  | -  | +    |

$V_{\beta}$  usage was analyzed in T-cell blasts after stimulation with each of the toxins shown. Most data are from B10.BR mice (H-2<sup>k</sup>). To analyze  $V_{\beta}$  elements that recognize IE-dependent superantigens IE<sup>-</sup> mice were used: B10.Q<sub>βBR</sub> mice for  $V_{\beta}17$  and B10.Q mice for  $V_{\beta}5$  and  $V_{\beta}11$ . Staining with  $V_{\beta}$ -specific monoclonal antibodies was used where possible, otherwise RNA was prepared from each of the blast populations and  $V_{\beta}$  expression established using a quantitative dot blot. Either unstimulated (staining) or concanavalin-A-stimulated T-cell blasts (RNA dot blots) were used as control cells. Pluses indicate an enrichment in  $V_{\beta}$  expression in the toxin-stimulated T-cell blasts compared with the control cells of twofold or better. n.d. indicates not determined.

Nude mice and neonatal mice, both of which lack mature functional T cells, are extremely resistant to SEB (data not shown), implicating T cells in the mode of action of SEB. However, the hypothesis we wished to test was that the various mechanisms that mice use to limit  $V_{\beta}$  expression may be a protective mechanism to limit the possible targets for these toxins in vivo. Therefore, we devised an experiment to test this idea more directly. We bred mice that combined a genetic deletion with the appropriate tolerizing  $V_{\beta}$ -specific self-superantigens to lower drastically the levels of the T cells bearing the  $V_{\beta}$  elements recognizing SEB. One strain, B10.BR(βBR) contributed a genetic deletion involving all members of the  $V_{\beta}8$  family. The other (CBA/J) contributed the self-superantigens deleting  $V_{\beta}3^{+}$  (Mls-2<sup>a</sup> or Mls-3<sup>a</sup>) and  $V_{\beta}7^{+}$  (unknown locus) T cells. (B10.BR[βBR] × CBA/J)F<sub>1</sub> mice were bred with B10.BR(βBR) mice and the progeny screened for the homozygous gene deletion and heterozygous but dominant superantigens. These mice occurred approximately with the predicted 1 in 8 frequency. On average, the sum of  $V_{\beta}(3 + 7 + 8)$  T cells in these mice was less than 6% compared with 32% in control B10.BR mice, which lack both the genetic deletion and the tolerizing self-superantigens. All mice in these experiments lacked  $V_{\beta}17^{+}$  T cells both because they expressed the IE-molecule of H-2<sup>k</sup> and in some cases because they bore the inactive  $V_{\beta}17$  gene. The experimental and control mice were injected with SEB, and their weight was followed over the next several days (Table 5). Whereas the control mice with a high level of SEB-reactive T cells showed a typical response with rapid weight loss, the mice specifically low in SEB-reactive T cells were protected against the toxin.

These results demonstrate that  $V_{\beta}$ -specific T-cell stimulation is important in the mode of action of these toxins in mice. The advantage to the bacteria of massive T-cell activation is not immediately apparent; however, our preliminary experiments have shown that after SEB administration mice temporarily produce a poor T-cell response to other antigens, which may be of some advantage to the bacteria producing these toxins.

### IMPLICATIONS

Our experiments establish that in mice the selective advantage of a large, diverse TCRαβ repertoire is not strong enough to weed out any of the many polymorphisms that limit  $V_{\beta}$  expression in laboratory and wild mice. Rather, they suggest an equilibrium in which mice exist in a state where all  $V_{\beta}$  elements are available to the species as a whole, but individuals express only a subset of the total. This implies some selective disadvantage to the expression of too many  $V_{\beta}$  elements in one individual. We suggest that this disadvantage lies in the susceptibility of mice to microbial toxins that rely on the massive stimulation of T cells bearing particular  $V_{\beta}$  elements for their toxicity. Our finding that mice constructed to have the right combination of  $V_{\beta}$  gene deletions and tolerizing  $V_{\beta}$ -specific self-superantigens are resistant to the appropriate toxin is consistent with this view.

Our results imply that the  $V_{\beta}$  repertoire of the mouse population is very flexible, capable of changing rapidly to accommodate the threat either from a new pathogen, the immune response to which may require a particular  $V_{\beta}$  element, or from a new microbial toxin in which case elimination of particular  $V_{\beta}$  elements may

Table 5. Deletion of T Cells Bearing Approximate  $V_{\beta}$  Elements Protects Mice from SEB

| Mouse  | T cells<br>bearing<br>$V_{\beta}3, 8, \text{ or } 7$<br>(%) | Weight loss following SEB (%) |              |             |
|--|---|-------------------------------|--------------|-------------|
|  |   | day 1                         | day 2        | day 3       |
| B10.BR   | 32  | 10.9<br>±0.7                  | 9.7<br>±1.0  | 5.3<br>±1.6 |
| (B10.Q[βBR] × CBA/J)F <sub>1</sub><br>× B10.Q(βBR) | 6   | 2.6<br>±1.4                   | -0.2<br>±1.1 | 2.6<br>±1.1 |

Mice were given 50 μg of SEB intraperitoneally on day 0.

be protective. The self-superantigens are a particularly efficient way of limiting expression of particular  $V_{\beta}$  elements although keeping them in reserve for rapid reexpression should the need arise. Suppression of a particular  $V_{\beta}$  is achieved without altering the  $V_{\beta}$  structural gene in mice heterozygous for the relevant superantigen. Mice expressing  $V_{\beta}$  appear in every mating between superantigen heterozygous mice. For example, to maintain the lack of  $V_{\beta}6$  expression in 90% of mice, a  $V_{\beta}$  haplotype with a  $V_{\beta}6$  gene deletion must occur at a frequency of 95% in the gene pool, and the reemergence of  $V_{\beta}6$  can only come from matings involving the 10% of mice already expressing  $V_{\beta}6$ . On the other hand, lack of  $V_{\beta}6$  expression can be maintained at the 90% level with a frequency of only 68% of the Mls-1<sup>a</sup> gene, and approximately 30% of matings, including those between two Mls-1<sup>a</sup> heterozygous mice each of which lacks  $V_{\beta}6$  expression, will yield at least some  $V_{\beta}6$ -expressing progeny.

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